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The potential role of exercise in chronic stress-related changes in AMPA receptor phenotype underlying synaptic plasticity

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[Purpose] Chronic stress can cause disturbances in synaptic plasticity, such as long-term potentiation, along with behavioral defects including memory deficits. One major mechanism sustaining synaptic plasticity involves the dynamics and contents of α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptors (AMPA) in the central nervous system. In particular, chronic stress-induced disruption of AMPARs includes its abnormal expression, trafficking, and calcium conductance at glutamatergic synapses, which contributes to synaptic plasticity at excitatory synapses. Exercise has the effect of promoting synaptic plasticity in neurons. However, the contribution of exercise to AMPAR behavior under chronic stressful maladaptation remains unclear.

[Methods] The present article reviews the information about the chronic stress-related synaptic plasticity and the role of exercise from the previous-published articles.

[Results] AMPAR-mediated synaptic transmission is an important for chronic stress-related changes of synaptic plasticity, and exercise may at least partly contribute to these episodes.

[Conclusion] The present article discusses the relationship between AMPARs and synaptic plasticity in chronic stress, as well as the potential role of exercise.

[Key words] exercise, memory, depression, synaptic plasticity, AMPAR

INTRODUCTION

Stress affects a variety of body systems including the neural, endocrine, immune, and digestive systems. Stress hormones, such as corticosterone, are regulated by the hypothalamic-pituitary-adrenal axis for alertness and adaptation in response to any demand and/or threat ^{1,2}. Sustained increases in corticosterone levels result in hippocampal atrophy, impaired long-term potentiation, and reduced neurogenesis, which produces aberrant synaptic plasticity and behavioral abnormalities ³⁻⁵. Chronic stress leads to diverse deteriorative consequences in the brain, which in turn impairs cognitive processes, such as learning and memory, and develops into emotion and mood-related illness such as depression ⁶⁻⁸.

Chronic stress-induced neuronal and behavioral abnormalities are deeply related to synaptic plasticity. Synaptic plasticity refers to the ability of synapses to strengthen or weaken over time in response to altered activity. Aberrant synaptic plasticity, including structural and functional plasticity, emerges under the maladaptive condition evoked by chronic stress ^{9,10}. Chronic stress-induced abnormalities in synaptic plasticity are modulated by corticosterone, neurotrophins, oxidative stress, and various neurotransmitters ^{8,11-13}. For example, chronic mild stress has been shown to reduce hippocampal transcription of hippocampal brain-derived natriuretic factor (BDNF), which reached basal levels in an isoform-specific manner by KCl-treated depolarization, suggesting that neurotrophins differentially regulate activity-dependent transcription of BDNF ¹⁴.

Glutamate, the major excitatory neurotransmitter released from presynaptic terminals, binds to specific receptors that are clustered in the postsynaptic membrane, which mediate the depolarizing signals in glutamatergic synapses. In particular, α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptors (AMPA) is a fast ligand-gated cation channel. AMPAR-dependent synaptic influx of cations, especially Ca²⁺, plays a critical role in synaptic plasticity ¹⁴⁻¹⁶. Diverse chronic stress leads to abnormal

synaptic function of AMPAR, including AMPAR-mediated excitation in the synapse ¹⁷⁻¹⁹.

Exercise has a beneficial effect on brain functions in both physiological and pathological states. Several studies have suggested that a variety of exercises can enhance hippocampal neurogenesis and neurotransmission underlying the synaptic plasticity related to cognition and mood under normal ^{20, 21} and chronic stress conditions ^{22, 23}. Thus, chronic stress-induced disturbances in synaptic plasticity can result in abnormal neuronal responses and behavioral defects, which can be, at least in part, overcome by exercise intervention. The present review discusses the literature pertaining to the relationship between chronic stress-induced abnormal AMPAR characteristics and synaptic plasticity, as well as the potential role of exercise.

The general mechanisms of synaptic plasticity

In general, synaptic plasticity refers to changes in synaptic strength depending on increases or decreases in activity. Synaptic plasticity represents a fundamental mechanism of enabling neurons to generate adaptive responses to stimuli for learning and memory. Subtypes of synaptic plasticity are classified into short-term, long-term, and homeostatic, so-called “synaptic scaling”.

Short-term synaptic plasticity refers to changes in synaptic efficacy over time, depending on the history of presynaptic activity within hundreds to thousands of microseconds. Short-term depression is produced by neurotransmitter(s) depletion during the synaptic signaling process at the axonal terminal of a pre-synaptic neuron, while short-term facilitation is mediated by Ca²⁺ influx into the axonal terminal according to spike production, which increases the probability of neurotransmitter(s) release and leads to structural changes (i.e., shape and density) in dendritic spines ²⁴.

Unlike short-term plasticity, long-term plasticity is primarily modulated by gene expression and protein synthesis. Long-term potentiation (LTP), and its counterpart long-term depression (LTD), are two forms of long-term plasticity. Changes at excitatory synapses are long-lasting (i.e., minutes or more). LTD is generated by a minimum level of postsynaptic depolarization as well as increases in the intracellular calcium concentration at the postsynaptic neuron. LTP is the increased synaptic response after the potentiation of prolonged electrical stimuli above the baseline response for hours or longer. Long-lasting synaptic stabilization is regulated by structural changes, including pre- and post-synaptic density, along with the increase in the postsynaptic density protein-95, which causes synaptic enlargement ²⁵.

Homeostatic plasticity refers to the ability of neurons to modify and self-adjust their excitability over a timescale of days. Homeostatic plasticity maintains the stability of

neuronal functions through coordinated plasticity among subcellular compartments, such as synapses versus the neurons, and cell bodies versus axons, unlike synapse-specific correlation-based plasticity mechanisms such as LTP and LTD ²⁶.

The role of the AMPAR in synaptic plasticity

Over the past decades, the molecular mechanisms underpinning synaptic plasticity have been extensively investigated in models of learning and memory. In particular, one of the major mechanisms involved in synaptic plasticity is the dynamics and activity of AMPA-type receptors for controlling plastic changes in the strength and connectivity of glutamatergic or excitatory synapses.

AMPA receptors are formed from the tetrameric assembly of subunits GluR1-4, which mediates the fast moment-to-moment transmission of excitatory signals on post-synapses. Endogenous forms of AMPAR primarily consist of GluR1/GluR2 or GluR2/GluR3 heteromers. Glutamatergic synapses that lack AMPAR current—known as “silent synapses”—are not able to achieve sufficient depolarization (excitation) despite containing functional N-methyl-D-aspartate (NMDA) receptors ²⁷. In addition to NMDA receptor-dependent Ca²⁺ influx, AMPAR-dependent synaptic Ca²⁺ influx is required for NMDA receptor-mediated LTP ^{14-15, 28}. Additionally, enhanced LTP expression has been observed in mice with genetic deletion of GluR2 ^{29, 30}. The phenotypic properties of long-term AMPARs, including synaptic recruitment and calcium permeability, are believed to play a critical role in NMDAR-dependent LTP. These AMPAR dynamics for synaptic plasticity are regulated by its biosynthesis, dendritic transport, exocytosis and endocytosis, through interaction with partner proteins and translational modifications.

The contents and trafficking of the AMPAR into the plasma membrane through endocytosis and exocytosis is the key regulator of plasticity at glutamatergic synapses ¹⁴.

This process is related to Hebbian and homeostatic plasticity, and is implicated in the interaction of several proteins. For example, the association of GluR2/3 with C-kinase 1 contributes to LTP, LTD, and homeostatic plasticity ^{31, 32}. The binding of GluR2 to N-ethylmaleimide-sensitive factor also contributes to synaptic incorporation through SNARE-mediated membrane fusion, in which the heteromeric GluR1/GluR2 receptor is recruited into the synaptic site by CaMKII activation ³³. Recent studies have reported that overexpression of neural precursor cell-expressed developmentally downregulated gene 4-1 (Nedd4-1), a member of the E3 ligase family, reduced the surface density of AMPARs through facilitated endocytosis and the accumulated internalization of GluR1 in the endosome ³⁴⁻³⁵. Downregulation of homeostatic scaling maintains internal ex-

citability via control of synaptic AMPARs content under sustained enhancement of synaptic activity by GABAA receptor antagonism or chronic increased neuronal activity³⁵⁻³⁶.

Synaptic insertion of AMPAR into the plasma membrane of excitatory neurons is regulated by protein kinases; PKA- and CaMKII-dependent GluR1 phosphorylation, produces or stabilizes more synapses, thereby controlling synaptic plasticity^{28, 37, 38}. As mentioned above, the characteristics of AMPAR behavior are an important determinant of synaptic plasticity.

Chronic stress and AMPARs under chronic stress-induced maladaptation, and the role of exercise

Chronic stress-induced elevation of glucocorticoid levels affects glutamate transmission and synaptic plasticity, thereby leading to abnormal behavior(s) such as cognitive impairment and depression³⁹. Although, exercise has long been known to improve synaptic plasticity, exercise-elicited AMPAR phenotype alteration in chronic stressful conditions has rarely been investigated. Mounting evidence has demonstrated how chronic stress induces disturbances in AMPAR-dependent synaptic plasticity. For example, chronic stress led to a reduction in AMPAR-dependent excitation of temporoammonic (TA)-CA1 path synapses and a decrease in AMPAR expression of hippocampal CA1¹⁷⁻¹⁸. Chronic restraint stress causes an alteration in AMPAR distribution and function, as well as an increase in neuronal excitatory drive on the basolateral amygdala⁴⁰⁻⁴¹. Our previous data revealed increased GluR1 content and PKA-directed GluR1 phosphorylation in the basolateral amygdala synapse, along with behavioral depression¹³.

Significant evidence supports the exercise-elicited improvement of synaptic plasticity. For example, acute and long-term exercise enhanced the hippocampal expression of synaptic plasticity-related genes, which includes synaptic remodeling-related genes such as synapsin I and synaptotagmin, as well as synaptic plasticity-promoting pathways such as CaMK II and BDNF [20]. Our previously published study demonstrated that 4-week treadmill running restored chronic stress-induced decreases in hippocampal BDNF expression in an AMPK-dependent manner, along with the reversal of memory impairment²². With regard to the AMPAR, voluntary exercise reversed the decreased field excitatory post-synaptic potential of Schaffer collateral-CA1 pathway concomitantly with enhanced GluR2, which is the less Ca²⁺-permeable AMPAR assembly, in a genetic rat model of depression²³. Furthermore, 4 weeks of voluntary wheel running—but not acute exercise—enhanced GluR1 and pGluR1 (Ser845) levels in the hippocampus⁴¹. In unpublished data, exercise exerted an ampa-like effect on chronic stress-induced failure of memory

consolidation and depression-like behaviors, indicated by rendering AMPAR Ca²⁺ permeable in the CA1 area of the hippocampus.

Apart from the hippocampus, repeated exercise alters the distribution of AMPAR subunits in diverse brain regions, evidenced by the alteration of AMPAR subunits depending on the duration of sensory-motor cortical area, cerebellum, and striatum⁴². In the mesolimbic reward pathway, which is closely associated with stress-response plasticity, 6-week running enhanced the expression of tyrosine hydroxylase messenger RNA in the ventral tegmental area and delta opioid receptor in the shell region of the nucleus accumbens⁴³, in which dopamine signaling alters AMPAR-mediated synaptic transmission or potentiation in the nucleus accumbens shell⁴⁴, suggesting that endogenous dopamine may affect AMPA receptor-mediated Ca²⁺ conductance.

As addressed above, chronic stress disrupts AMPAR-mediated synaptic plasticity in some limbic structures, such as the hippocampus, thereby leading to behavioral abnormalities such as cognitive- and mood-related illness. In contrast, exercise may help cope with chronic stress-induced aberrant synaptic plasticity by the incorporation of calcium-permeable AMPAR into the synapse, thereby improving stress-related consequences.

Prospective

Chronic stress-induced defects in behavior(s), such as impairment of memory processes and mood-related disorders, are closely linked to synaptic plasticity. This phenomenon has, at least in part, been attributed to the characteristics of AMPARs such as calcium conductance and trafficking. To date, however, the relationship between these electrophysiological and molecular events and exercise has rarely been explored. To clarify this issue, there are several promising prospects. First, we need to investigate what molecules are able to regulate synaptic AMPAR expression, and to determine what signals or partners control AMPAR trafficking in “exercised environments”. Second, to clarify which aspects of an exercise program, including intensity, type, and duration, efficiently affects and/or effects AMPAR-dependent alteration of AMPAR properties.

REFERENCES

1. Kwon DH, Kim BS, Chang H, Kim YI, Jo SA, Leem YH. Exercise ameliorates cognition impairment due to restraint stress-induced oxidative insult and reduced BDNF level. *Biochem Biophys Res Commun*. 2013;434:245-51.
2. Magarinos AM, McEwen BS. Stress-induced atrophy of apical dendrites of hippocampal CA3c neurons: involvement of glucocorticoid secretion and excitatory amino acid receptors. *Neuroscience*. 1995;69:89-98.
3. Liston C, Gan WB. Glucocorticoids are critical regulators of dendritic spine development and plasticity in vivo. *Proc Natl*

- Acad Sci USA*. 2011;108:16074–9.
4. Pavlides C, Ogawa S, Kimura A, McEwen BS. Role of adrenal steroid mineralocorticoid and glucocorticoid receptors in long term potentiation in the CA1 field of hippocampal slices. *Brain Res*. 1996;738:229–35.
 5. Han TK, Lee JK, Leem YH. Chronic exercise prevents repeated restraint stress-provoked enhancement of immobility in forced swimming test in ovariectomized mice. *Metab Brain Dis*. 2015;30:711–8.
 6. Cryan, JF, Holmes, A. The ascent of mouse: advances in modeling human depression and anxiety. *Nat Rev Drug Discov*. 2005;4:775–90.
 7. Berton, O, McClung, CA, DiLeone, RJ, Krishnan, V, Renthal, W, Russo, SJ, et al. Essential Role of BDNF in the Mesolimbic Dopamine Pathway in Social Defeat Stress. *Science*. 2006;311:864–8.
 8. Sandi C, Pinelo-Nava MT. Stress and memory: behavioral effects and neurobiological mechanisms. *Neural Plast*. 2007;2007:78970.
 9. Christian KM, Miracle AD, Wellman CL, Nakazawa K. Chronic stress-induced hippocampal dendritic retraction requires CA3 NMDA receptors. *Neuroscience*. 2011;174:26–36.
 10. Yang Q, Zhu G, Liu D, Ju JG, Liao ZH, Xiao YX, et al. Extrasynaptic NMDA receptor dependent long-term potentiation of hippocampal CA1 pyramidal neurons. *Sci Rep*. 2017;7:3045.
 11. McGaugh JL, Roozendaal B. Role of adrenal stress hormones in forming lasting memories in the brain. *Curr Opin Neurobiol*. 2002;12:205–10.
 12. Yamada K, Nabeshima T. Brain-derived neurotrophic factor/TrkB signaling in memory processes. *J Pharmacol Sci*. 2003;91:267–70.
 13. Yi ES, Oh S, Lee JK, Leem YH. Chronic stress-induced dendritic reorganization and abundance of synaptosomal PKA-dependent CP-AMPA receptor in the basolateral amygdala in a mouse model of depression. *Biochem Biophys Res Commun*. 2017;486:671–8.
 14. Molteni R, Rossetti AC, Savino E, Racagni G, Calabrese F. Chronic Mild Stress Modulates Activity-Dependent Transcription of BDNF in Rat Hippocampal Slices. *Neural Plast*. 2016;2016:2592319.
 15. Huganir RL, Nicoll RA. AMPARs and synaptic plasticity: the last 25 years. *Neuron*. 2013;8:704–17.
 16. Plant K, Pelkey KA, Bortolotto ZA, Morita D, Terashima A, McBain CJ. Transient incorporation of native GluR2-lacking AMPA receptors during hippocampal long-term potentiation. *Nat Neurosci*. 2006;9:602–4.
 17. Makino H, Malinow R. AMPA receptor incorporation into synapses during LTP: the role of lateral movement and exocytosis. *Neuron*. 2009;64:381–90.
 18. Kallarackal AJ, Kvarita MD, Cammarata E, Jaber L, Cai X, Bailey AM, et al. Chronic stress induces a selective decrease in AMPA receptor-mediated synaptic excitation at hippocampal temporoammonic-CA1 synapses. *J Neurosci*. 2013;33:15669–74.
 19. Schmidt MV, Trümbach D, Weber P, Wagner K, Scharf SH, Liebl C, et al. Individual stress vulnerability is predicted by short-term memory and AMPA receptor subunit ratio in the hippocampus. *J Neurosci*. 2010;30:16949–58.
 20. Molteni R, Ying Z, Gomez-Pinilla F. Differential effects of acute and chronic exercise on plasticity-related genes in the rat hippocampus revealed by microarray. *Eur J Neurosci*. 2002;16:1107–16.
 21. Dietrich MO, Mantese CE, Porciuncular LO, Ghisleni G, Vinade L, Souza DO, et al. Exercise affects glutamate receptors in postsynaptic dendrites from cortical mice brain. *Brain Res*. 2005;1065:20–5.
 22. Kim DM, Leem YH. Chronic stress-induced memory deficits are reversed by regular exercise via AMPK-mediated BDNF induction. *Neuroscience*. 2016;324:271–85.
 23. Gomez-Galan M, Femenia T, Aberg E, Graae L, Van Eeckhaut A, Smolders I, et al. Running opposes the effects of social isolation on synaptic plasticity and transmission in a rat model of depression. *PLoS One*. 2016;11:0165071.
 24. Lamprecht R, LeDoux J. Structural plasticity and memory. *Nat Rev Neurosci*. 2004;5:45–54.
 25. Meyer, D, Bonhoeffer T, Scheuss V. "Balance and Stability of Synaptic Structures during Synaptic Plasticity". *Neuron*. 2014;82:430–43.
 26. Turrigiano G. Homeostatic Synaptic Plasticity: Local and Global Mechanisms for Stabilizing Neuronal Function. 2012. *Cold Spring Harb Perspect Biol*. 2012;4:005736.
 27. Rao VR, Finkbeiner S. NMDA and AMPA receptors: old channels, new tricks. *Trends Neurosci*. 2007;30:284–91.
 28. Esteban JA, Shi S, Wilson C, Nuriya M, Huganir RL, Malinow R. PKA phosphorylation of AMPA receptor subunits controls synaptic trafficking underlying plasticity. *Nat Neurosci*. 2003;6:136–43.
 29. Asrar S, Zhou Z, Ren W, Jia Z. Ca(2+) permeable AMPA receptor induced long-term potentiation require PI3/MAP kinase but Ca/CaM-dependent kinase II. *PLoS One*. 2009;4:e4339.
 30. Wiltgen BJ, Royle GA, Gray EE, Abdipranoto A, Thangthaeng N, Jacobs N, Saab F, Tonegawa S, Heinemann SF, O'Dell TJ, Fanselow MS, Vissel B. A role for calcium-permeable AMPA receptors in synaptic plasticity and learning. *PLoS One*. 2010;5:e12818.
 31. Citri A, Bhattacharyya S, Ma C, Morishita W, Fang S, Rizo J, et al. Calcium binding to PICK1 is essential for the intracellular retention of AMPA receptors underlying long-term depression. *J Neurosci*. 2001;30:16437–52.
 32. Widagdo J, Fang H, Jang SE, Anggono V. PACSIN1 regulates the dynamics of AMPA receptor trafficking. *Sci Rep*. 2016;6:31070.
 33. Shi SH, Hayashi Y, Esteban JA, Malinow R. Subunit-specific rules governing AMPA receptor trafficking to synapses in hippocampal pyramidal neurons. *Cell*. 2001;105:331–43.
 34. Schwarz LA, Hall BJ, Patrick GN. Activity-dependent ubiquitination of GluA1 mediates a distinct AMPA receptor endocytosis and sorting pathway. *J Neurosci*. 2010;30:16718–29.
 35. Scudder SL, Goo MS, Cartier AE, Molteni A, Schwarz LA, Wright R. Synaptic strength is bidirectionally controlled by opposing activity-dependent regulation of Nedd4-1 and USP8. *J Neurosci*. 2014;34:16637–49.
 36. Jewett KA, Zhu J, Tsai NP. The tumor suppressor p53 guides GluA1 homeostasis through Nedd4-2 during chronic elevation of neuronal activity. *J Neurochem*. 2015;135:226–33.
 37. Bolshkov Vy, Golan H, Kandel ER, Siegelbaum SA. Re-

- cruitment of new sites of synaptic transmission during the cAMP-dependent late phase of LTP at CA3-Ca1 synapses in the hippocampus. *Neuron*. 1997;19:635-51.
38. Sheng M, Lee SH. AMPA receptor trafficking and the control of synaptic transmission. *Cell*. 2011;105:825-28.
 39. Chaouloff F, Groc L. Temporal modulation of hippocampal excitatory transmission by corticosteroids and stress. *Front Neuroendocrinol*. 2011;32:25-42.
 40. Hubert GW, Li C, Rainnie DG, Muly EC. Effects of stress on AMPA receptor distribution and function in the basolateral amygdala. *Brain Struct Funct*. 2014;219:1169-79.
 41. Venezia AC, Quinlan E, Roth SM. A single bout of exercise increases hippocampal BDNF: Influence of chronic exercise and noradrenaline. *Genes, Brain and Behavior*. 2017;16:800-11.
 42. Real CC, Ferreira AFB, Hernandez MS, Britto LRG, Pires RS. Exercise-induced plasticity of AMPA-type glutamate receptor subunits in the rat brain. *Brain Res*. 2010;1363:63-71.
 43. Greenwood BN1, Foley TE, Le TV, Strong PV, Loughridge AB, Day HE, et al. Long-term voluntary wheel running is rewarding and produces plasticity in the mesolimbic reward pathway. *Behav Brain Res*. 2011;217:354-62.
 44. Ingebretson AE, Hearing MC, Huffington ED, Thomas MJ. Endogenous dopamine and endocannabinoid signaling mediate cocaine-induced reversal of AMPAR synaptic potentiation in the nucleus accumbens shell. *Neuropharmacology*. 2017;131:154-65.